



Nanoscale mapping of the directional flow patterns at liquid-solid interfaces

Piantanida, L., Farokh Payam, A., Zhong, J., & Voitchovsky, K. (2020). Nanoscale mapping of the directional flow patterns at liquid-solid interfaces. *Physical review applied*, 13(6), [064003].
<https://doi.org/10.1103/PhysRevApplied.13.064003>

[Link to publication record in Ulster University Research Portal](#)

Published in:
Physical review applied

Publication Status:
Published (in print/issue): 01/06/2020

DOI:
[10.1103/PhysRevApplied.13.064003](https://doi.org/10.1103/PhysRevApplied.13.064003)

Document Version
Author Accepted version

General rights
Copyright for the publications made accessible via Ulster University's Research Portal is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The Research Portal is Ulster University's institutional repository that provides access to Ulster's research outputs. Every effort has been made to ensure that content in the Research Portal does not infringe any person's rights, or applicable UK laws. If you discover content in the Research Portal that you believe breaches copyright or violates any law, please contact pure-support@ulster.ac.uk.

Nanoscale mapping of the directional flow patterns at liquid-solid interfaces

Luca Piantanida^{1‡}, Amir F. Payam^{1†} Jing Zhong² and Kislou Voitchovsky^{1*}

¹ Department of Physics, Durham University, Durham, United Kingdom

² School of Civil Engineering, Harbin Institute of Technology, Harbin, China (zhongjing@hit.edu.cn)

[†] Present address: School of Engineering, Ulster University, Belfast, United Kingdom (a.farokh-payam@ulster.ac.uk)

[‡] Present address: Micron School of Materials Science & Engineering, Boise State University, Boise, USA (lucapiantanida@boisestate.edu)

* correspondence: kislou.voitchovsky@durham.ac.uk

Abstract

The nanoscale behavior of liquid molecules and solutes along the interface with solids controls many processes such as molecular exchanges, wetting, electrochemistry, nanofluidics, biomolecular function and lubrication. Experimentally, several techniques can explore the equilibrium molecular arrangement of liquids near the surface of immersed solids but quantifying the nanoscale flow patterns naturally adopted by this interfacial liquid remains a considerable challenge.

Here we describe a novel approach based on atomic force microscopy and able to quantify the flow direction preferentially adopted by liquids along interfaces with nanoscale precision. The approach, called vortex dissipation microscopy (VDM), uses high-frequency directional oscillations to derive local flow information around each location of the interface probed. VDM effectively derives nanoscale flow charts of the interfacial liquid parallel to a solid and can operate over a broad range of soft and hard interfaces. To illustrate its capabilities, we quantify the dynamics of aqueous solutions containing KCl or MgCl₂ along the surface of a same graphene oxide flake. We show that dissolved K⁺ ions can move evenly in all direction along the interface whereas Mg²⁺ ions tend to move in registry with the underlying lattice due to enthalpic effects. The results, provide *in-situ* nanoscale insights into the ion-specific sieving properties of graphene oxide membranes.

I. Introduction

In bulk liquids molecules diffuse randomly in all directions unless an external perturbation is imposed; no long-range molecular order is present. This picture breaks down when liquid molecules meet a wall. At the interface with immersed solids, liquid molecules and dissolved solutes tend to move more slowly and often adopt a more ordered arrangement compared to the when in the bulk [1-3]. This is due to a loss of configurational entropy and the molecules interacting with the solid's surface. This so-called interfacial liquid is ubiquitous in nature and in technology where it underpins numerous phenomena. Examples include protein function [4,5], molecular diffusion at interfaces during self-assembly [6], the functionalization of surfaces [7], wetting [8], lubrication [9], nanofluidics [10], electrochemistry and heterogeneous catalysis [11] and the functioning of energy cells [12] to name but a few.

Understanding the properties and behavior of interfacial liquid is therefore of paramount importance to the scientists and technologists alike. The task is however challenging because the interfacial liquid typically spans only a few molecular layers ($\sim < 2$ nm) near the surface of the solid. On regular interfaces such as single crystals, the most accurate measurements typically rely on averaging over a large number of interfacial molecules to increase the signal-to-noise ratio of measurements [3]. Averaging approaches can be problematic for irregular interfaces or systems where local contextual information is important. This is the case for many biological and electrochemical systems where the efficiency of interfacial processes hinge on particular nanoscale topographic or chemical features at a given location of the solid's surface. These difficulties can in principle be overcome by using local techniques such as atomic force microscopy (AFM) where a nanoscale probe is used to explore the interface. Recent advances in the field of AFM have enabled significant improvements with sub-nanometer spatial resolution being now routine [13-15], 3-dimensional mapping of the liquid density in the vicinity of solids [16,17], high-accuracy mapping of the interfacial rheology and the viscoelastic properties of soft biological samples [18,19], and high-speed measurements [20-22].

While these developments have enabled a wealth of new research, it is usually the unperturbed equilibrium molecular arrangement of the interfacial liquid that is being probed. No information about the local liquid dynamics or natural convective flows are available on that scale. This is a serious limitation of the technique because interfacial processes are dynamical in essence and often rely on the flow motion of liquid molecules near the solid's surface to proceed.

One possible way forward is to exploit the fact that AFM physically probes the interface with a moving nanoscale tip. As the tip moves across or through the interfaces and pushes the liquid molecules, it experiences a resistance related to the ability of the molecules to flow along the solid's surface [23]. Physically, this is quantified by the so-called slip length of the liquid at the interface, the distance beneath the solid's surface where the lateral velocity component of the liquid molecules would vanish, based on the extrapolated flow profile [24,25]. In aqueous solutions, the energy needed to displace the liquid molecules located immediately under the tip is directly related to the local slip-length [23], itself determined by the local affinity of the liquid for the solid [26]. This relationship quantitatively links the ability of liquid molecules to flow along the interface with the measurable energy needed to move the tip through the interface, therefore offering an opportunity to quantify nanoscale interfacial flow patterns. Practically, this entails the ability for the tip to probe all the possible flow directions around a given location and accurately quantify the energy dissipation associated with each direction. This task is currently not possible with existing AFM systems.

In this study we develop a novel approach to derive directional and quantitative information about the ability of liquids to flow laterally at solid-liquid interfaces with nanometer resolution. The method, based on a commercial AFM augmented with ultrafast nanoactuators, effectively maps the local flow patterns naturally occurring at the interface, similarly to the maps of maritime currents in navigation. In order to illustrate the method's capability, we use it to investigate the flow of aqueous solutions containing potassium chloride (KCl) and magnesium chloride (MgCl_2) at the interface with sheets of graphene oxide (GO). The two types of cations (K^+ and Mg^{2+}) are well-known to behave very differently when sieved through membranes composed of stacked GO sheets [27,28]. Water molecules can easily permeate through the membrane but cation-specific permeation has been reported [29,30] explained by differences in the ions' hydration structures when moving at the interface with GO [31]. Mapping the flow patterns forming at the surface of GO in the presence of each type of salt provides novel molecular-level insights into the mechanisms enabling ion-specific sieving by GO membranes.

II. Methods

A. Samples preparation

The imaging solutions were made using ultrapure water (18.2 M Ω , Merck-Millipore, Watford, UK) and chemicals purchased from Sigma Aldrich (Gillingham, UK) and used without further purification.

Grade V mica, purchased from SPI supplies (West Chester, PA, USA), was freshly cleaved before adding a drop (100 μ l) of solution and starting the experiment immediately after.

Bovine lens membranes were purified and deposited onto freshly cleaved mica following a protocol described elsewhere in detail [32]. In short, 10 μ l of lens membrane fragments were dissolved in a 30 μ l of a solution composed of 25 mM MgCl₂, 150 mM KCl and 10 mM Tris at pH 7.4. The resulting solution was deposited onto freshly cleaved mica and incubated for 15 minutes. Subsequently, the sample was rinsed gently 6 times with a solution containing 150 mM KCl and 10 mM Tris at pH 7.4 to remove loosely attached membrane fragments. More solution was then added to conduct the experiment.

GO was prepared from natural graphite (SigmaAldrich, <45 μ m) flakes by a modified Hummers' method as described in refs [33,34]. As-prepared GO being in gel state, it was subjected to thorough dialysis against ultrapure water to completely remove residual salts and acids. This resulted into a 10 mg/ml aqueous dispersion of GO flakes in ultrapure water. The surface of GO contains is highly charge in aqueous solutions, mainly due to its carboxyl, hydroxyl and alkoxyl groups which ensure its perfect dispersion in aqueous solutions. Near neutral pH and in conditions close to the present study, Zeta potential measurements reveal a negatively charged surface with values close to -30 mV [35-37]. For the experiments, the GO dispersion was diluted 10 times in 10 mM KCl reaching a final concentration of 1 mg/ml. A droplet (50 μ l) of this dispersion was placed on top of a freshly cleaved HOPG (SPI supplies) and incubated for 40 minutes at room temperature. The sample was then slowly rinsed 6 times with more 10 mM KCl solution to remove loosely adsorbed GO flakes and prevented the deposition of flakes on the AFM cantilever. More clean solution was then added to conduct the experiment.

B. AFM and VDM imaging

All the experiments were conducted on a commercial Cypher ES AFM (Asylum Research, Oxford Instruments, Santa Barbara, CA, USA) and with the tip and cantilever fully immersed into the liquid. The vertical oscillation of the tip was controlled by photothermal excitation (Blue Drive) and the experiments were conducted at 25.0 ± 0.1 °C (controlled). Before imaging a small droplet of the desired solution was used to pre-wet the AFM cantilever and holder so

as to avoid the formation of air bubbles. All experiments were conducted with the feedback on the tip oscillation amplitude (amplitude modulation mode). The imaging setpoint was optimized for VDM, typically requiring lower setpoint values than for standard AFM [38]. In order to allow for direct comparability between measurements on GO, the experiments were carried out in 10 mM KCl and 10 mM MgCl₂ over an identical location (same GO flake), with the same tip and same imaging conditions. To avoid possible cross-contamination, the experiment was first conducted in KCl and subsequently in MgCl₂, using extensive rinsing in-between.

III. Results

A. The Vortex Dissipation Microscope

The basic principles of the novel microscope are presented in Fig. 1. The idea is to combine the high-resolution imaging capabilities of dynamic mode AFM –a vertically vibrating tip at a set interface location– with the directional information available from shear force microscopy measurements –a laterally moving tip at a set distance from the solid–. When operated with amplitudes comparable to the size of the interface (typically < 1-2nm), both approaches have been shown to probe local liquid slippage at the interface [14,23]. In the former, the liquid molecules are pushed indiscriminately as the tip squeezes out the interfacial liquid [14] while in the latter the direction of motion of the liquid molecules is defined, but at the cost of lateral resolution [23]. In order to retain the high-resolution capabilities of dynamic mode AFM while simultaneously gaining the directional insights from shear force microscopy, a combined vertical and lateral oscillatory motion is applied to the tip. Practically, this is achieved by operating the AFM in a standard dynamic mode (here amplitude modulation with a tip vertical oscillation frequency ν_{tip}) and imposing a lateral oscillation to the sample with respect to the tip, along a controllable direction and with a velocity comparable to that of the vertical tip motion (Fig. 1a).

In order to explore all the possible directions around a given location of the sample, the lateral motion is formed by two modulated oscillations along perpendicular X and Y directions (Fig. 1b). The faster oscillation (frequency ν_{sweep}) imposes a rapid lateral sweep of the tip along a given direction. The slower oscillation (frequency ν_{vortex}) progressively changes the direction of the sweep, systematically exploring all the possible lateral directions over its period (Fig.

1c). The resulting motion of the stage is a rapid directional sweep which direction periodically rotates so as to complete a full cycle over a period τ_{vortex} . To reflect the vortex-like scanning motion of the stage, the method is hereafter called *vortex dissipation microscopy* (VDM).

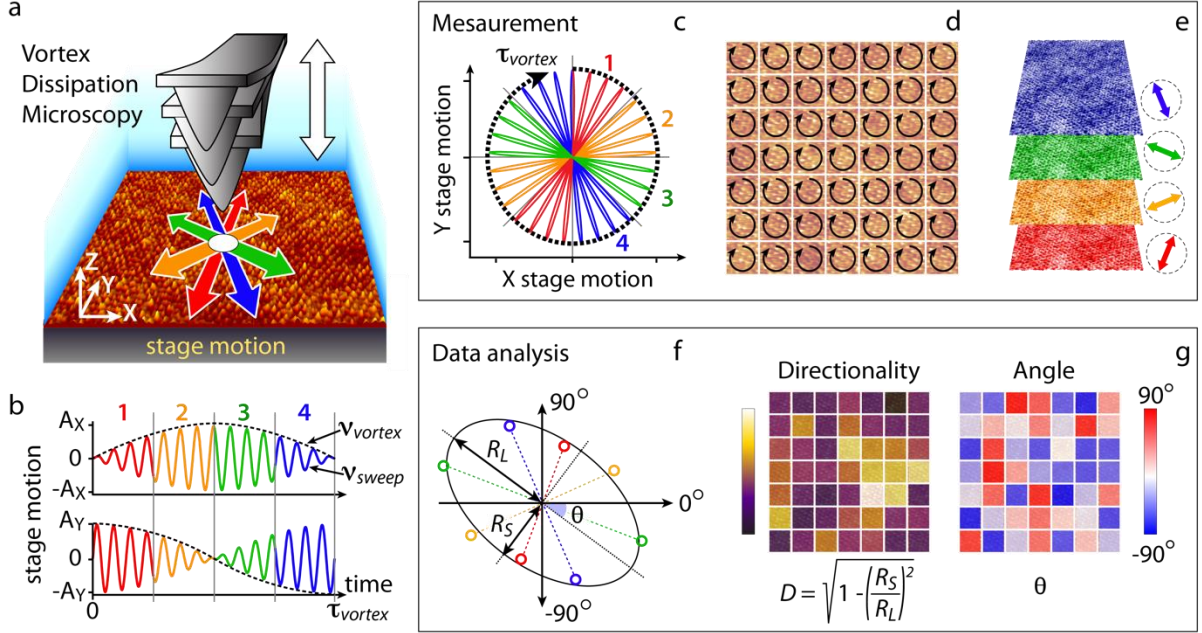


FIG. 1 (color online) Working principles of the vortex dissipation microscopy. High-resolution dynamical mode AFM is combined with shear force microscopy to create a convoluted vertical and lateral oscillation of the tip with respect to the sample (a). The lateral oscillation (b) combines a rapid sinusoidal motion at frequency v_{sweep} modulated by a slower frequency v_{vortex} . The resulting stage motion creates rapid lateral sweeps with slowly changing direction, probing every lateral direction around a given sample point (whole revolution or vortex) in τ_{vortex} (c). By sequencing the duration of a full vortex in a set of regular time intervals (here 4) and averaging the AFM measurement parameters (amplitude, phase, frequency) over each interval, directional information can be associated with each AFM parameter for the interface location considered. The process can be repeated for each imaging pixel of the interface investigated (d), creating multiple directional charts of the lateral liquid flow (e). The data analysis is conducted pixel by pixel, plotting in polar coordinate the directional data associated with a given pixel (f). Here no phase synchronization is kept between the vertical and vortex oscillations so a given direction cannot distinguish which way (forward or backward) is preferred. Two symmetrical points are hence plotted for each direction: the angle represent the direction and the distance to origin the magnitude of the associated parameter. The polar plot is subsequently fitted with an ellipse (larger radius R_L , a shorter radius R_S and an orientation angle θ). The degree of directionality is given by the ellipse eccentricity $D = \sqrt{1 - \left(\frac{R_S}{R_L}\right)^2}$ with $0 < D < 1$ and $D = 0$ indicating no preferential direction. A value of D and θ is obtained for each pixel (g).

In order to quantify the energy necessary to move the liquid in a particular direction, the energy dissipated by the tip can be calculated from the standard AFM operating parameters [39-41]

directly available through the AFM controller (amplitude, phase and frequency of the vertical oscillation), while the stage sweeps along that exact direction. The magnitude of the vertical and lateral components of the tip average velocity have to be kept comparable to ensure a meaningful directional measurement. A full directional readout is then achieved at any given location of the interface by successively quantifying the energy dissipation associated with each sweep direction over a full vortex. To do this, the vortex period is divided in smaller time windows, each associated with an average direction (time intervals 1-4 in Fig. 1b, c). This strategy allows for a reasonably fast vortex acquisition while ensuring each direction is probed by multiple sweep oscillations (typically > 10) for reliable measurements. The process is repeated for each pixel of the interface to map the ease with which the liquid flows in every lateral direction (Fig. 1d). This results in a series of maps, each quantifying the energy dissipation associated with a given direction for each image pixel (Fig. 1e).

The data analysis is then conducted pixel by pixel: all the directional information associated with a given pixel is plotted in polar coordinates, with for each direction the angle and distance to the origin representing respectively the flow direction and the ease for the liquid to flow in that direction (see section 1 of the Supplemental Material for details). Subsequent fitting with an ellipse determines the dominating flow direction at the pixel considered (Fig. 1f). This information is given by the ellipse eccentricity, called here directionality D . If the flow is equivalent in all directions, then the ellipse is a circle and $D = 0$. In contrast, if a given direction dominates, then $D > 0$ and the associated ellipse angle θ quantifies this dominating direction (Fig. 1g). It should be pointed out that for values of D close to zero, the angle is not defined regardless of the fit result.

B. Hardware development

The VDM method is in principle compatible with any modern AFM, provided that a lateral vortex modulation can be applied and that the data acquisition is properly synchronized so as to enable the directional analysis described above. Here this is achieved by a purposely built piece of hardware, the ‘vortex scanner’, made to sit directly on top of the normal AFM scanner with the sample placed on top of the vortex scanner (Fig. 2a). In this way, both the standard AFM scanner and the vortex scanner can function simultaneously, with their respective motions adding up. For VDM measurements, the vortex scanner only needs to operate at two frequencies (ν_{sweep} and ν_{vortex}), with a continuous modulated sinusoidal motion in both x and y directions, and at small amplitudes (< 1 nm) to allow for high spatial resolution. It was

therefore designed and built with two goals: (i) small size to fit within with most commercial AFMs and (ii) stable operation at high frequencies. The result is a small rectangular scanner of size of 12 mm \times 12 mm \times 2 mm (Fig. 2b-e) with a natural resonance frequency close to 35 kHz (Fig. 2f). Details of the design and calibration are discussed in the section 2 of the Supplemental Material.

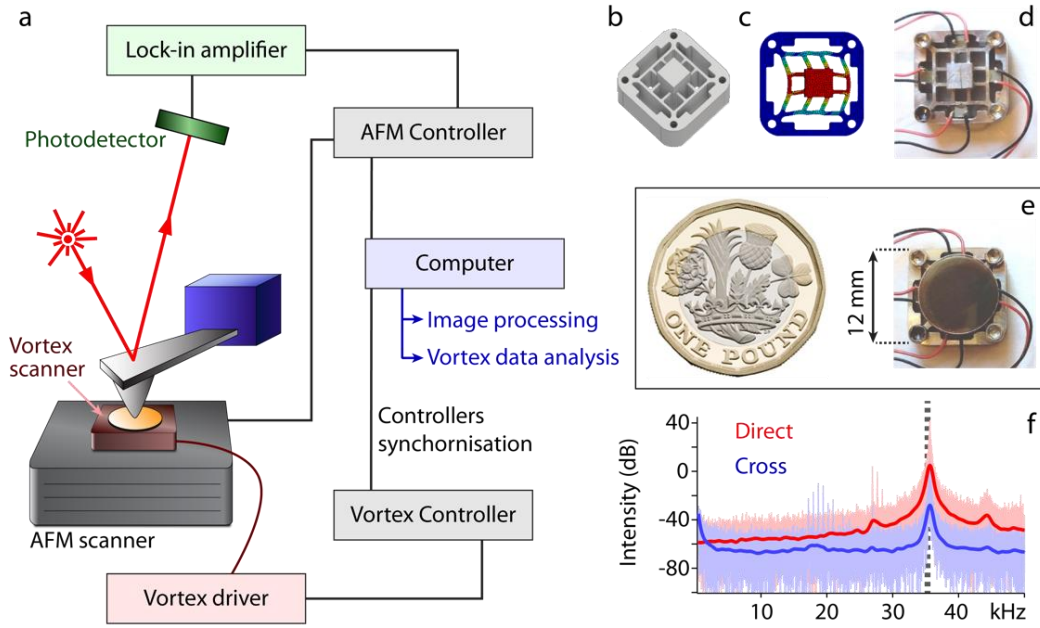


FIG. 2 (color online) Schematic representation of the VDM hardware assembly. The vortex scanner supports the sample and is mounted atop the AFM scanner (a). The electronics controlling the AFM and the VDM are synchronized to ensure completion of exactly one vortex per pixel and appropriate data acquisition. The vortex scanner design (b) has been optimized using finite element simulation of its operation (c). The scanner realization includes X and Y driving piezo elements, here with counter-piezo to quantify its motion performance (d). The sample is mounted on the central part (e). The scanner size is 12 mm \times 12 mm \times 2 mm, smaller than a British £1 coin (to scale). When the scanner is driven with a single piezo at 4 V and 1 Hz (f), the counter-piezo and cross-piezo can be used to measure respectively the direct (red curve) and cross-talk (blue) mechanical performance. A resonance can be seen around 35 kHz, very close to the finite elements predicted value (dashed vertical line).

C. High-resolution capabilities and no-flow reference

The main motivation for developing the VDM is to gain the ability to map interfacial flow patterns with nanoscale lateral precision. We therefore tested first the impact of the VDM operation on the high-resolution capabilities of the setup in various conditions to ensure that local nanoscale measurements are possible. The mechanical properties of the interface examined are expected to significantly influence the resolution achieved [42,43] and testing

was therefore conducted in aqueous solutions at the interface with a hard mica surface and a soft aquaporin 0 crystal extracted from cow lens membranes [32,44]. Furthermore since mica exhibits a no-slip boundary condition in water [23,45], no directionality information should be observed with VDM. representative results are presented in Fig. 3, showing the standard topography and phase information available in amplitude modulation AFM together with the directionality and orientation (angle) data obtained from the VDM operation.

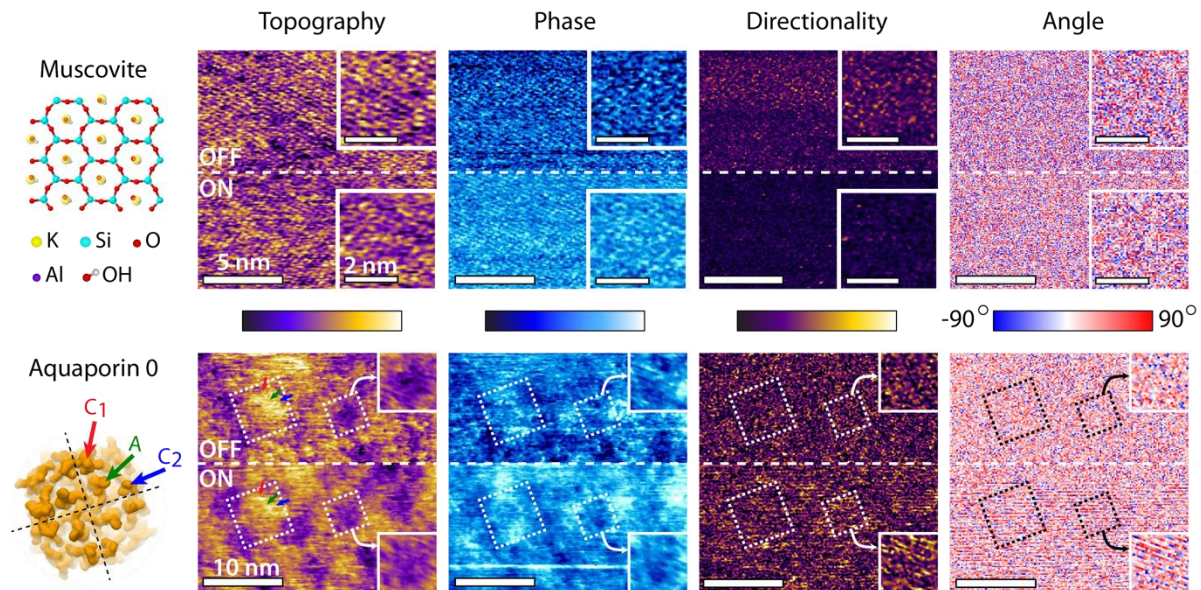


FIG. 3 (color online) Representative examples of high-resolution VDM operation. Relatively hard (muscovite mica, top) and soft (aquaporin 0, bottom) solids are investigated in aqueous solutions. Both sets of images are acquired with the AFM scanning top to bottom and the vortex operation switched ON mid-image (dashed white line) for direct comparison. Insets represent magnified portions of each image with the vortex OFF (upper) and ON (lower). A protein tetramer and an inter-tetramer region are highlighted on the aquaporin lattice (dashed squares). The mica is immersed in a 10 mM KCl solution and the aquaporin in Tris 10 mM, KCl 150 mM buffer solution (pH 7.4). The data on mica was acquired using a Scanasyst FLUID+ cantilever (Bruker, Coventry, UK) with a nominal stiffness $k \sim 0.7$, $v_{vertical} = 332$ kHz in solution (2nd eigenmode) and the vortex scanner operating with $v_{sweep} = 71.98$ KHz and $v_{vortex} = 512$ Hz. On aquaporin, the data was acquired with an Arrow UHF cantilever (Nanoworld, Neuchatel, Switzerland) with stiffness $k \sim 2$ N/m, $v_{vertical} = 303$ kHz in solution and the vortex scanner operating with $v_{sweep} = 50.176$ KHz and $v_{vortex} = 512$ Hz. The color scale bars represent height variations of 0.2 nm (mica) and 1.2 nm (aquaporin), phase variations of 20° (mica) and 10° (aquaporin), and a directionality varying between $0.01 < D < 0.45$ (mica) and $0.16 < D < 0.32$ (aquaporin).

The muscovite-water interface can be resolved with atomic-level resolution when the vortex scanner is switched ON (Fig. 3 upper part), revealing limited effect on the imaging quality.

This is made possible by the relatively small sweeping amplitude of the vortex (here ~ 0.3 nm peak to peak based on off-resonance calibration, see Supplemental Fig. S1). Generally, a compromise is needed between the spatial resolution achieved and deriving meaningful directional information. The former requires the sweep amplitude to be as small as possible while the latter needs for comparable vertical and lateral tip velocities with respect to the sample. This implies for v_{sweep} to be as large as possible, often requiring operation at frequencies higher than the vortex scanner resonance to obtain meaningful results. In practice we found a range of frequencies just above the scanner resonance (40-50kHz) where acceptable resolution can be achieved for horizontal amplitudes of up to ~ 2 nm (peak to peak), almost 10 times larger than the typical vertical oscillation amplitudes. This allows to compensate for that fact that $v_{tip} \sim 10 v_{sweep}$ while ensuring suitable sampling of the AFM observables. On mica, Fig. 3 shows some random fluctuations in directionality when the VDM is OFF, immediately disappearing to $D \sim 0$ when the VDM is turned ON. This is expected due to the fact that nanoscale lateral flow is negligible at the mica-water interface [23,45] (null slip length) and the VDM averages out any random fluctuations. The associated angle information is hence meaningless.

Interestingly, resolution on the aquaporin-solution interface improves when the VDM is switched ON, showing less distorted proteins and the distinctive crystal pattern in topography and phase, and the inter-tetramer space (smaller dashed square) appearing darker in the phase as previously reported [32]. The relatively harsh imaging conditions used when the VDM is OFF come from the stiff cantilever used here, but when the VDM is switched ON the lateral tip velocity enables the tip to ‘glide’ on the sample’s solvation layers[46], thereby preserving both sample and resolution. The directionality exhibits a faint relative increase over the inter-tetramer region (compared to VDM OFF) with an associated angle distribution slightly more polarized between extreme values. However, the small sampling region and the lack of proper flow calibration does not allow for any discussion beyond these qualitative observations.

We note that an ideal measurement scenario would have $v_{tip} = v_{sweep}$ and the vertical and horizontal motions synchronized in phase so as to enable not only directional information, but also differentiating between forward and backward motions of the tip in any given direction. While this is in principle possible, it could not be achieved with the current setup due to the lateral sweep stimulating the tip vertical oscillation in a highly directional manner when v_{sweep} approaches the cantilever resonance frequency.

D. Calibration of the VDM measurements

The results shown in Fig. 3 demonstrate that VMD can operate with high resolution over a variety of interfaces, but a full calibration of the system is necessary to derive any meaningful quantitative data. This includes not only information from the sample, but also from the measuring tip such as the impact of the tip geometry, chemistry and positioning (e.g. tilt angle) all of which may not be symmetrical in all lateral directions. To achieve this task, we investigated the interface between an aqueous solution and sheets of graphene oxide (GO) deposited onto an atomically flat, hydrophobic graphite (HOPG) substrate. HOPG exhibits a large slip length [23] rendering high-resolution with in amplitude modulation challenging [14]. Here, this can be used as a reference since the flow of water is expected to be identical all directions based on AFM observations, at least within the resolution of the VDM. In contrast, GO is hydrophilic and its ion-specific sieving capabilities [29,30] suggests the presence of directional effects in an aqueous solution. To exacerbate possible effects, the experiment is conducted in a solution of MgCl_2 since Mg^{2+} ions are sieved out of GO filters [27], indicating strong interfacial hydration effects on flow. Since both HOPG and GO can be made atomically flat, topographical effects on the VDM measurements can be excluded. The results and their analysis are detailed in Fig. 4.

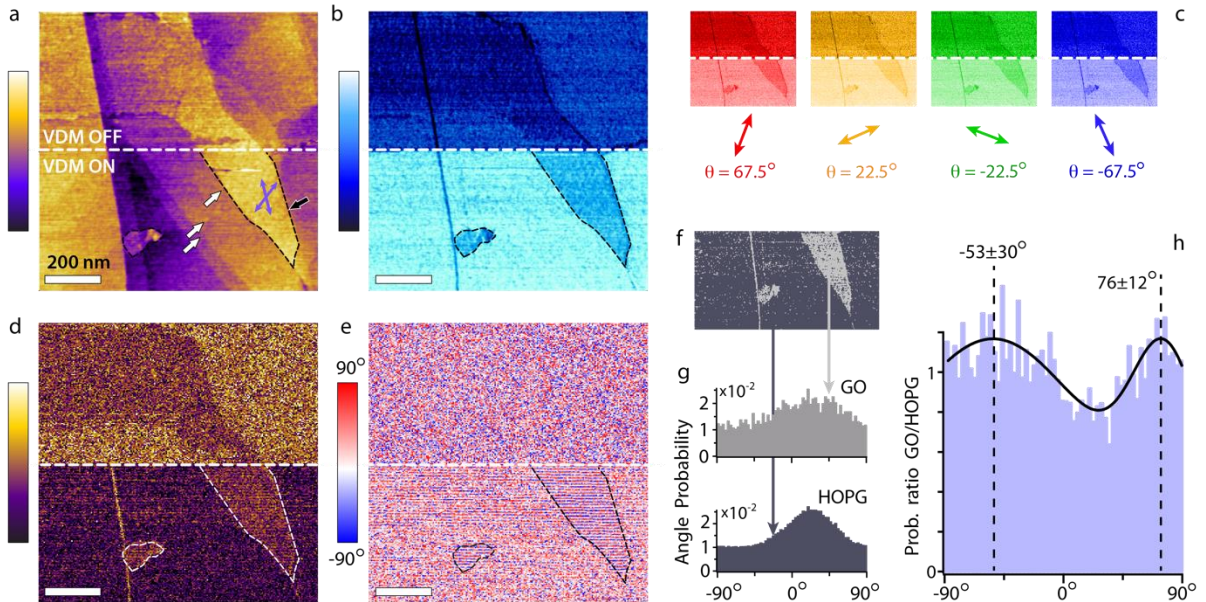


FIG. 4 (color online) VDM analysis of the interface between a GO flake and a 10 mM MgCl_2 solution. As for Fig. 3, the vortex scanner is switched ON halfway through the AFM scan so that the top part of all images

(VDM OFF) serves as reference. Topography (a) and phase images (b) show GO flakes (black dashed outline) on top of the HOPG surface. HOPG atomic steps are indicated with the white arrows and the edge of the GO flake with a black arrow in (a). Interestingly, the phase contrast between GO and HOPG is enhanced by the VDM with a darker phase over the GO. This confirms a high dissipation over GO and hence lower slippage or ease for the solution to flow. Using the VDM acquisition and analysis strategy described in Fig. 1, directional flow maps are acquired (c) and combined into directionality (d) and angle (e) images. GO flakes exhibit a higher directionality than HOPG and a lower (bluer) angle probability. A mask based on the phase contrast (f) is used to sample the angle distributions over HOPG and GO regions (g). The angle distribution over HOPG shows one clear maximum around 30° , presumably induced by tip effects such as vertical tilt and local chemistry. The distribution broadens and shifts towards lower angles over GO. Normalizing the GO distribution by the HOPG distribution provides a result free of tip effects (h). Two maxima are visible at $\sim 120^\circ$ from each other, obtained by fitting the normalized distribution with a sum of two Gaussians. The corresponding directions are indicated by purple arrows in (a), suggesting an alignment with the GO flake edge and its crystalline orientation. The data was acquired using a Scanasyt FLUID+ cantilever with $\nu_{vertical} = 305$ kHz in solution (2nd eigenmode) and the vortex scanner operating with $\nu_{sweep} = 40.96$ KHz and $\nu_{vortex} = 512$ Hz. The color scale bars represent height variations of 3 nm (a), phase variations of 30° (b), and a directionality varying between $0.15 < D < 0.45$ (d).

Due to atomic steps of the HOPG, it is not possible to distinguish the GO and the HOPG from the topographic information alone (Fig. 4a). However, the phase exhibits a clear contrast between both materials (Fig. 4b). This is expected since the phase is linked to the interfacial energy dissipation and hence sensitive to the local flow dynamics. Consistently, a lower phase (higher dissipation) is seen over the more hydrophilic GO. The directionality is also sensitive to the two materials: in the absence of vortex (Fig. 4d up), a marginally higher directionality is seen on HOPG compared to GO reflecting the higher lateral flow on HOPG, albeit in random directions due to local fluctuations. When the VDM is switched ON, the measurement becomes controlled and no directionality is any longer visible on HOPG, except at step edges (bright vertical line in the lower part of Fig. 4d). This apparent step directionality is however an artefact due to topographic effects and to the finite speed of the AFM feedback that induces local changes in the scanning conditions. Directionality changes at HOPG step edges were therefore ignored. In contrast, a clear and consistent directionality increase can be seen over the whole of the GO flake. The flow angle appears relatively uniform on HOPG with a bias toward positive (red) values. This bias is less marked on GO flakes. To better quantify the differences in angle between GO and HOPG, we used the phase information (Fig. 4b) to create a mask that objectively distinguishes GO and HOPG and calculated angle probability distributions on both regions (Fig 4f-g). The angle distribution exhibits a clear maximum on HOPG despite no topographical effects, pointing to tip effects instead. The measuring tip is bound to affect the

measurement by the fact that it is not perfect with some asymmetry in shape, chemistry, hydration and potentially also in its positioning with a likely tilt of the tip axis with respect to the interface's perpendicular direction. The measurement on HOPG therefore serves as a calibration to quantify this effect and exclude them from further measurements. Here this is done for GO by dividing the angle distribution obtained on GO by that on HOPG (Fig. 4h). Two distinct maxima appear at $\sim 120^\circ$ from each other (purple arrows in (a)) and aligned with the edge of the GO flake (black arrow). This clearly suggests a correlation between the preferential flow directions and the crystalline orientation of the GO. Significantly, the preferred directions are not well aligned with the HOPG's atomic steps (white arrows in (a)), reinforcing the interpretation that GO dominates the flow pattern.

E. Nanoscale flow patterns in different ionic solutions

To further confirm the VDM results, the same experiment was conducted in a 10 mM KCl solution, with the same AFM tip, in the same operating conditions, and over the same sample location. To avoid possible cross-contamination, the experiment was first conducted in KCl and subsequently in MgCl_2 with the latter presented in Fig. 3. A comparative result for both ionic solutions is given in Fig. 5.

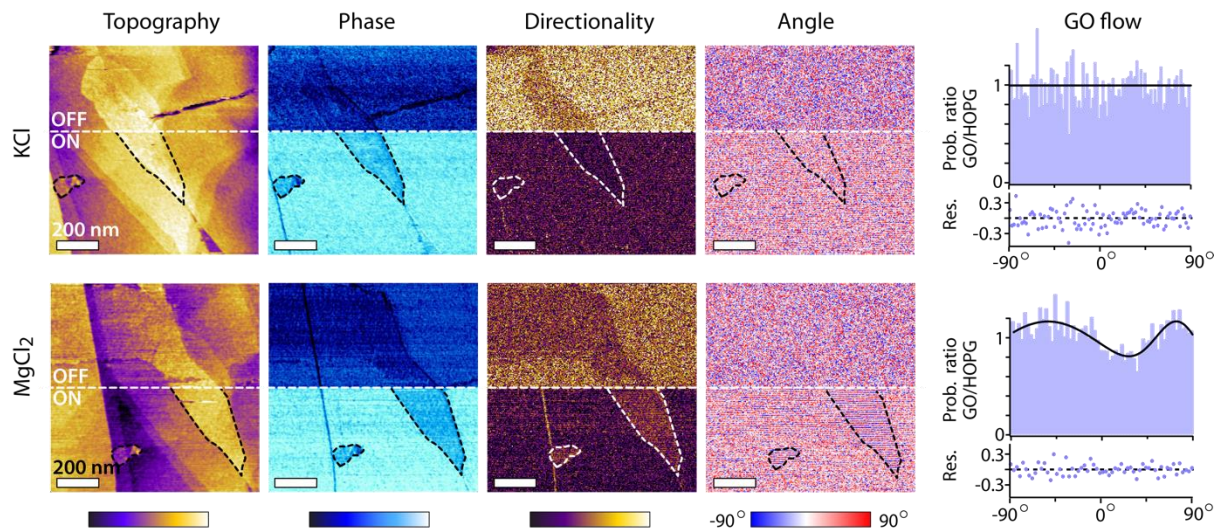


FIG. 5 (color online) Comparative VDM results for a same GO flake in a solution of KCl and MgCl_2 . The same sample location is investigated in both cases, in identical operating conditions and with the same tip. The results were acquired first in 10 mM KCl and subsequently in 10 mM MgCl_2 after extensively rinsing the sample with the MgCl_2 solution. The phase contrast between GO and HOPG is consistent and similar in magnitude in

both solutions, but no clear directionality contrast can be seen in KCl. Similarly, no preferential angle can be identified in the KCl solution. The angle distribution is fitted with a horizontal line in KCl and a double Gaussian in MgCl_2 , with the fit residuals given in both case. The data is presented as in Fig. 4 and is identical for MgCl_2 . The color scale bars are identical for both solutions and represent height variations of 3 nm (topography), 30° (phase), and $0.15 < D < 0.45$ (directionality).

Standard amplitude modulation AFM gives almost identical results in KCl and in MgCl_2 , but the VDM analysis shows significant differences in interfacial flow when Mg^{2+} ions are present in the solution. In KCl, no change in directionality can be seen between HOPG and GO, a result confirmed by the absence of any preferred angle once the tip effects have been discounted. However, preferential flow directions in registry with the GO lattice emerge in presence of Mg^{2+} ions. To understand the origin of these ion-specific differences, it is worth considering the hydrodynamic radii of the two cations with 3.31 Å for K^+ and 4.28 Å for Mg^{2+} [27,47]. Previous studies have shown the hydrodynamic radius of the cations present in solution to play a key role on their intercalation and percolation between GO flakes and determine the sieving performance of GO membranes [27,29,30]. The permeation rate is significantly larger for K^+ than for Mg^{2+} [27,28] with a total permeation cut-off for ions with hydrodynamic radii larger than ~ 4.5 Å [29]. Since GO membranes are typically composed of random stacks of GO flakes, reliable measurements of the permeation rates are challenging and results can be contradictory depending on the membrane preparation [48-50]. Generally, the permeation is explained in terms of geometrical constraints whereby larger ions and molecules simply cannot pass in the limited space available between adjacent GO sheets [30,51-53]. Electrostatic interactions between the moving ions and the GO's carbonyl groups have also been invoked to explain the swelling of GO stacks in the presence of MgCl_2 [31].

The present results suggest that the motion of metal ions at the surface of GO can be understood as a combination of restructuring dynamics of the water molecules between the ions' shell and the GO's hydrophilic groups, and direct electrostatic interactions between ions and the GO surface. The effect of hydration water on the dynamics of the ions is expected from previous studies with GO [29]. Water molecules have been shown to move at supra-diffusive velocities between GO sheets by primarily traveling on hydrophobic paths and avoiding GO's hydrophilic groups and flowing [54]. Adding hydrated ions to the system can partly be explained by the same picture whereby larger ions can no longer travel along these hydrophobic paths without interacting with GO's hydrophilic groups or shedding part of their hydration water to fit between GO sheets [29,30].

In this framework, the size dependence on the hydrated ions' radius can be explained with a combination of enthalpic and entropic free energy costs. Larger hydrated ions are usually more densely charged and experience stronger electrostatic interactions with the negatively charged GO surface. Moving along the interface therefore incur higher enthalpic costs for larger ions which are more likely to remain in registry with the underlying GO lattice. The effect is further exacerbated by the low dielectric constant of nano-confined water [55]. The entropy costs associated with the ions moving along the interface originate from the restructuring of water molecules in the ion's and GO's hydration shell. Size-related effects are likely to depend on the degree of confinement [56] and are hence less straightforward to predict than enthalpic contributions.

Similar experiments have been conducted with AFM to investigate the behavior of metal cations at the interface with mica in aqueous solutions [9,57]. Mica surface are atomically flat over large regions and exhibits a hexagonal lattice with a periodicity close to double that of graphene. In the present experimental conditions, mica is negatively charged with a surface potential of the order of -60 mV to -80 mV [58], about twice the value of GO [36]. When minimally confined (as is the case here), metal cations diffusing at the surface of mica show little dependence on temperature [57], confirming enthalpic effects to dominate. This is to be expected considering the larger surface potential. However, strong confinement with an AFM tip during shear experiments, the hydrogen bond network formed by the interfacial water molecules dominate the cation's mobility, with divalent cations moving more easily owing to their ability to better disrupt the local hydrogen bond order [9]. While on experiments conducted on mica cannot be directly transposed to the present case with mica being almost defect-free and able to stabilize interfacial water molecules in an ice-like arrangement [57,59], the fact that the opposite trend is observed here on GO suggests that enthalpic effects involving direct electrostatic interactions between the ions and the charged groups of the GO surface dominate. This is also supported by the fact that densely charged Mg^{2+} ions tend to move more easily when in registry with the underlying crystalline lattice.

Overall, the VDM results are consistent with the well-known poor permeability of typical GO membranes to Mg^{2+} ions and the variability between different permeability measurements [27,29,48]: the existence of a clear preferential direction for ions moving between two GO sheets hinges on the sheets having their respective lattice in registry. This is unlikely to be the case for most GO membranes unless created so by design.

F. General discussion and conclusion

The present study describes how AFM operation in liquid augmented with small lateral oscillations can be used to map the nanoscale flow patterns naturally adopted by the liquid parallel to the surface of immersed solids. The method is compatible with most existing AFMs and effectively exploits the boundary friction of the interfacial liquid when pushed in a given direction to quantitatively deduce the ease for the liquid to move in the direction considered. The VDM method builds on the fact that AFM measurements conducted within the interfacial liquid are highly sensitive to the boundary flow through the measured energy dissipation [14,23,42]. In order to derive meaningful flow information, most of the tip apex ($> 20 \text{ nm}^2$) is likely to contribute to the measured dissipation, but atomic-level resolution is still possible thanks to localized asperities [14,60] or stable hydration sites [61] located on the tip apex. Here, the validity of the VDM approach is demonstrated on reference samples such as mica and HOPG where no measurable directionality is expected. On mica, the boundary slip is known to be zero [23], hence preventing lateral flow in any direction. On HOPG, the boundary slip is relatively large, but AFM cannot identify any specific feature and hence preferred direction when operated as in the present study. Consistently, VDM measures zero directionality on mica, and a small directionality on HOPG, the latter being due to expected tip effects that can subsequently be discounted.

Our results demonstrate that the approach is also compatible with high-resolution imaging and can be exploited to gain insights into the nanoscale fluid dynamics at interfaces. Quantitative measurements taken on GO membranes illustrate directly and *in-situ* the ion-dependent directional flow effects. While such effects are known to occur, the present results provide novel insights into the underpinning molecular mechanisms and suggest that electrostatic interactions through the hydration water may dominate the process over equilibrium steric effects. Arguably, the images (Fig. 4-5) are relatively low resolution when compared to those presented in Fig. 3, but this is due to the emphasis being placed on obtaining quantitative directional information and on reproducibility (see Supplemental Fig. S2 for a repeat experiment with another tip). Higher resolution is also possible on the HOPG/GO systems when relaxing the requirement of using a same tip over a same sample location, with details of the interface resolved down to the nanometer level (Supplemental Fig. S3).

Beyond the new measurements rendered possible here with the VDM, there is still scope for improvement and exploration of the method's possibilities. The vortex scanner, for example, can be improved for higher resonance and lower noise operation, also operating the acquisition off-resonance [22] to avoid cross-talk between lateral and vertical oscillations. In terms of the

measurement process itself, the VDM can be exploited to modulate and enhance the spatial resolution and explore the dynamics of different solvation layers at interfaces, as illustrated in Fig. 3 over aquaporin. This is because the interfacial liquid itself plays an important role in AFM resolution at solid-liquid interfaces [14,43]. On soft samples such as biomembranes this can be further exploited to preserve the sample by playing with the viscoelastic properties of the interfacial water which then acts as a lubrication, energy absorbing layer [62]. Future work will explore the full capabilities of VDM, in particular on biological samples where membrane fluidity [63], interfacial molecular diffusion and biomolecular dynamics [4,5,32] are closely related to the properties of interfacial liquid.

Acknowledgements

The authors acknowledge funding from the Biotechnology and Biological Sciences Research Council (grant BB/M024830/1) (LP, AFP, and KV), and the Royal Society (grant RG2014R2) (KV).

Authors Contributions

LP conducted all the experiments. AFP developed the vortex scanner and implemented the VDM software. JZ synthesized the GO flakes. KV designed the project and coordinated its implementation. LP and KV wrote the paper with input from AFP and JZ.

References

- [1] W. Kaplan and Y. Kauffmann, Structural order in Liquids induced by Interfaces with Crystals, *Annu. Rev. Mat. Res.* **36**, 1 (2006).
- [2] C. Yu, G. Evmenenko, A. Richter, A. Datta, J. Kmetko, and P. Dutta, Order in molecular liquids near solid-liquid interfaces, *Appl. Surf. Sci.* **182**, 231 (2001).
- [3] P. Fenter and N. Sturchio, Mineral-water interfacial structures revealed by synchrotron X-ray scattering, *Prog. Surf. Sci.* **77**, 171 (2004).
- [4] H. Frauenfelder, G. Chena, J. Berendzena, P. W. Fenimore, H. Janssonb, B. H. McMahon, I. R. Stroe, J. Swenson, and R. D. Young, A unified model of protein dynamics, *Proc. Natl. Acad. Sci. U.S.A.* **106**, 5129 (2009).
- [5] M. Grossman, B. Born, M. Heyden, D. Tworowski, G. B. Fields, I. Sagi, and M. Havenith, Correlated structural kinetics and retarded solvent dynamics at the metalloprotease active site, *Nat. Struct. Mol. Biol.* **18**, 1102 (2011).

- [6] K. Voitchovsky, D. Giofrè, J. José Segura, F. Stellacci, and M. Ceriotti, Thermally-nucleated self-assembly of water and alcohol into stable structures at hydrophobic interfaces, *Nat. Commun.* **7**, 13064 (2016).
- [7] J. Love, L. Estroff, J. Kriebel, R. Nuzzo, and G. Whitesides, Self-assembled monolayers of thiolates on metals as a form of nanotechnology, *Chem. Rev.* **105**, 1103 (2005).
- [8] L. Gao and T. McCarthy, Wetting 101°, *Langmuir* **25**, 14105 (2009).
- [9] C. Cafolla and K. Voitchovsky, Lubricating properties of single metal ions at interfaces, *Nanoscale* **10**, 11831 (2018).
- [10] U. F. Keyser, B. N. Koeleman, S. van Dorp, D. Krapf, R. M. M. Smeets, S. G. Lemay, N. H. Dekker, and C. Dekker, Direct force measurements on DNA in a solid-state nanopore, *Nat. Phys.* **2**, 473 (2006).
- [11] M. Kühne, F. Börrnert, S. Fecher, M. Ghorbani-Asl, J. Biskupek, D. Samuelis, A. V. Krasheninnikov, U. Kaiser, and J. H. Smet, Reversible superdense ordering of lithium between two graphene sheets, *Nature* **564**, 234 (2018).
- [12] K. Voitchovsky, N. Ashari-Astani, I. Tavernelli, N. Tetreault, U. Rothlisberger, F. Stellacci, M. Grätzel, and H. A. Harms, In Situ Mapping of the Molecular Arrangement of Amphiphilic Dye Molecules at the TiO₂ Surface of Dye-Sensitized Solar Cells, *ACS Appl. Mater. Interf.* **7**, 10834 (2015).
- [13] T. Fukuma, M. Higgins, and S. Jarvis, Direct Imaging of Lipid-Ion Network Formation under Physiological Conditions by Frequency Modulation Atomic Force Microscopy, *Phys. Rev. Lett.* **98**, 106101 (2007).
- [14] K. Voitchovsky, J. J. Kuna, S. A. Contera, E. Tosatti, and F. Stellacci, Direct mapping of the solid-liquid adhesion energy with subnanometre resolution, *Nat. Nanotechnol.* **5**, 401 (2010).
- [15] W. Trewby, J. Faraudo, and K. Voitchovsky, Long-lived ionic nano-domains can modulate the stiffness of soft interfaces, *Nanoscale* **284**, 173 (2019).
- [16] T. Fukuma, Y. Ueda, S. Yoshioka, and H. Asakawa, Atomic-Scale Distribution of Water Molecules at the Mica-Water Interface Visualized by Three-Dimensional Scanning Force Microscopy, *Phys. Rev. Lett.* **104**, 016101 (2010).
- [17] D. Martin-Jimenez, E. Chacon, P. Tarazona, and R. Garcia, Atomically resolved three-dimensional structures of electrolyte aqueous solutions near a solid surface, *Nat. Commun.* **7**, 12164 (2016).
- [18] X. Xiong, S. Guo, Z. Xu, P. Sheng, and P. Tong, Development of an atomic-force-microscope-based hanging-fiber rheometer for interfacial microrheology, *Phys. Rev. E* **80**, 061604 (2009).
- [19] D. Guan, E. Charlaix, R. Z. Qi, and P. Tong, Noncontact Viscoelastic Imaging of Living Cells Using a Long-Needle Atomic Force Microscope with Dual-Frequency Modulation, *Phys. Rev. Appl.* **8**, 044010 (2017).
- [20] T. Uchihashi, N. Kodera, and T. Ando, Guide to video recording of structure dynamics and dynamic processes of proteins by high-speed atomic force microscopy, *Nat. Protoc.* **7**, 1193 (2012).
- [21] T. Uchihashi, R. Iino, T. Ando, and H. Noji, High-Speed Atomic Force Microscopy Reveals Rotary Catalysis of Rotorless F₁-ATPase, *Science* **333**, 755 (2011).

- [22] A. P. Nievergelt, N. Banterle, S. H. Andany, P. Gönczy, and G. E. Fantner, High-speed photothermal off-resonance atomic force microscopy reveals assembly routes of centriolar scaffold protein SAS-6, *Nat. Nanotechnol.* **13**, 696 (2018).
- [23] D. Ortiz-Young, H. C. Chiu, S. Kim, K. Voïtchovsky, and E. Riedo, The interplay between apparent viscosity and wettability in nanoconfined water, *Nat. Commun.* **4**, 2482 (2013).
- [24] C.-L. M. H. Navier, Sur les lois de l'équilibre et du mouvement des corps solides élastiques, *Mém. Acad. Roy. Sci.*, France 839 (1821).
- [25] L. R. Bocquet and J.-L. Barrat, Flow boundary conditions from nano- to micro-scales, *Soft Matter* **3**, 685 (2007).
- [26] D. M. Huang, C. Sendner, D. Horinek, R. R. Netz, and L. Bocquet, Water Slippage versus Contact Angle: A Quasiuniversal Relationship, *Phys. Rev. Lett.* **101**, 226101 (2008).
- [27] J. Abraham, K. S. Vasu, C. D. Williams, K. Gopinadhan, Y. Su, C. T. Cherian, J. Dix, E. Prestat, S. J. Haigh, I. V. Grigorieva, P. Carbone, A. K. Geim, and R. R. Nair, Tunable sieving of ions using graphene oxide membranes, *Nat. Nanotechnol.* **12**, 546 (2017).
- [28] J.-H. Song, H.-W. Yu, M.-H. Ham, and I. S. Kim, Tunable Ion Sieving of Graphene Membranes through the Control of Nitrogen-Bonding Configuration, *Nano Lett.* **18**, 5506 (2018).
- [29] R. K. Joshi, P. Carbone, F. C. Wang, V. G. Kravets, Y. Su, I. V. Grigorieva, H. A. Wu, A. K. Geim, and R. R. Nair, Precise and Ultrafast Molecular Sieving Through Graphene Oxide Membranes, *Science* **343**, 752 (2014).
- [30] B. Mi, Graphene Oxide Membranes for Ionic and Molecular Sieving, *Science* **343**, 740 (2014).
- [31] Y. Mo, X. Zhao, and Y.-X. Shen, Cation-dependent structural instability of graphene oxide membranes and its effect on membrane separation performance, *Desalination* **399**, 40 (2016).
- [32] M. Ricci, R. A. Quinlan, and K. Voïtchovsky, Sub-nanometre mapping of the aquaporin–water interface using multifrequency atomic force microscopy, *Soft Matter* **13**, 187 (2017).
- [33] N. I. Kovtyukhova, P. J. Ollivier, B. R. Martin, T. E. Mallouk, S. A. Chizhik, E. V. Buzaneva, and A. D. Gorchinskiy, Layer-by-Layer Assembly of Ultrathin Composite Films from Micron-Sized Graphite Oxide Sheets and Polycations, *Chem. Mater.* **11**, 771 (1999).
- [34] W. Hummers and R. Offeman, Preparation of Graphitic Oxide, *J. Am. Chem. Soc.* **80**, 1339 (1958).
- [35] J. Zhong, W. Sun, Q. Wei, X. Qian, H.-M. Cheng, and W. Ren, Efficient and scalable synthesis of highly aligned and compact two-dimensional nanosheet films with record performances, *Nat. Commun.* **9**, 1 (2018).
- [36] G. Xu, J. Zhong, and X. Shi, Influence of graphene oxide in a chemically activated fly ash, *Fuel* **226**, 644 (2018).

- [37] Y. Yu, J. Zhong, W. Sun, R. Kumar, and N. Koratkar, Solid-State Hybrid Fibrous Supercapacitors Produced by Dead-End Tube Membrane Ultrafiltration, *Adv. Funct. Mater.* **27**, 1606461 (2017).
- [38] E. J. Miller, W. Trewby, A. Farokh Payam, L. Piantanida, C. Cafolla, and K. Voïtchovsky, Sub-nanometer Resolution Imaging with Amplitude-modulation Atomic Force Microscopy in Liquid, *J. Vis. Exp.* **118**, e54924 (2016).
- [39] H. Söngen, R. Bechstein, and A. Kühnle, Quantitative atomic force microscopy, *J. Phys.: Condens. Matter* **29**, 274001 (2017).
- [40] J. Cleveland, B. Anczykowski, A. Schmid, and V. Elings, Energy dissipation in tapping-mode atomic force microscopy, *Appl. Phys. Lett.* **72**, 2613 (1998).
- [41] J. Sader and S. Jarvis, Coupling of conservative and dissipative forces in frequency-modulation atomic force microscopy, *Phys. Rev. B* **74**, 195424 (2006).
- [42] K. Voïtchovsky, Anharmonicity, solvation forces, and resolution in atomic force microscopy at the solid-liquid interface, *Phys. Rev. E* **88**, 022407 (2013).
- [43] K. Voïtchovsky and M. Ricci, High-resolution imaging of solvation structures with amplitude-modulation atomic force microscopy, *Proc. SPIE*, edited by W. J. Parak, K. Yamamoto, and M. Osinski (SPIE, San Francisco, California, USA, 2012), pp. 82320O–8.
- [44] N. Buzhynskyy, P. Sens, F. Behar-Cohen, and S. Scheuring, Eye lens membrane junctional microdomains: a comparison between healthy and pathological cases, *New J. Phys.* **13**, 085016 (2011).
- [45] T.-D. Li and E. Riedo, Nonlinear Viscoelastic Dynamics of Nanoconfined Wetting Liquids, *Phys. Rev. Lett.* **100**, 106102 (2008).
- [46] A. Humphris, M. Miles, and J. Hobbs, A mechanical microscope: High-speed atomic force microscopy, *Appl. Phys. Lett.* **86**, 034106 (2005).
- [47] J. Mähler and I. Persson, A Study of the Hydration of the Alkali Metal Ions in Aqueous Solution, *Inorg. Chem.* **51**, 425 (2011).
- [48] A. Morelos-Gomez, R. Cruz-Silva, H. Muramatsu, J. Ortiz-Medina, T. Araki, T. Fukuyo, S. Tejima, K. Takeuchi, T. Hayashi, M. Terrones, and M. Endo, Effective NaCl and dye rejection of hybrid graphene oxide/graphene layered membranes, *Nat. Nanotechnol.* **12**, 1083 (2017).
- [49] Z. Jia, W. Shi, Y. Wang, and J. Wang, Dicarboxylic acids crosslinked graphene oxide membranes for salt solution permeation, *Coll. Surf., A* **494**, 101 (2016).
- [50] J. Wang, L. Zhao, D. Wei, W. Wu, J. Zhang, and X. Cheng, Effects of Intercalated Molecules in Graphene Oxide on the Interlayer Channels for Anhydrous Proton Conduction, *Ind. Eng. Chem. Res.* **55**, 11931 (2016).
- [51] L. Huang, Y. Li, Q. Zhou, W. Yuan, and G. Shi, Graphene Oxide Membranes with Tunable Semipermeability in Organic Solvents, *Adv. Mater.* **27**, 3797 (2015).
- [52] A. Klechikov, J. Yu, D. Thomas, T. Sharifi, and A. V. Talyzin, Structure of graphene oxide membranes in solvents and solutions, *Nanoscale* **7**, 15374 (2015).
- [53] A. V. Talyzin, T. Hausmaninger, S. You, and T. Szabó, The structure of graphene oxide membranes in liquid water, ethanol and water–ethanol mixtures, *Nanoscale* **6**, 272 (2014).

- [54] D. W. Boukhvalov, M. I. Katsnelson, and Y.-W. Son, Origin of Anomalous Water Permeation through Graphene Oxide Membrane, *Nano Lett.* **13**, 3930 (2013).
- [55] L. Fumagalli, A. Esfandiar, R. Fabregas, S. Hu, P. Ares, A. Janardanan, Q. Yang, B. Radha, T. Taniguchi, K. Watanabe, G. Gomila, K. S. Novoselov, and A. K. Geim, Anomalous low dielectric constant of confined water, *Science* **360**, 1339 (2018).
- [56] D. Argyris, D. R. Cole, and A. Striolo, Ion-Specific Effects under Confinement: The Role of Interfacial Water, *ACS Nano* **4**, 2035 (2012).
- [57] M. Ricci, W. Trewby, C. Cafolla, and K. Voitchovsky, Direct observation of the dynamics of single metal ions at the interface with solids in aqueous solutions, *Sci. Rep.* **7**, 43234 (2017).
- [58] Z. Adamczyk, M. Zaucha, and M. Zembala, Zeta Potential of Mica Covered by Colloid Particles: A Streaming Potential Study, *Langmuir* **26**, 9368 (2010).
- [59] Y. Leng and P. T. Cummings, Fluidity of Hydration Layers Nanoconfined between Mica Surfaces, *Phys. Rev. Lett.* **94**, 026101 (2005).
- [60] D. J. Müller, D. Fotiadis, S. Scheuring, S. A. Müller, and A. Engel, Electrostatically Balanced Subnanometer Imaging of Biological Specimens by Atomic Force Microscope, *Biophys. J.* **76**, 1101 (1999).
- [61] S. M. R. Akrami, H. Nakayachi, T. Watanabe-Nakayama, H. Asakawa, and T. Fukuma, Significant improvements in stability and reproducibility of atomic-scale atomic force microscopy in liquid, *Nanotechnology* **25**, 455701 (2014).
- [62] K. Voitchovsky, S. A. Contera, and J. F. Ryan, Lateral coupling and cooperative dynamics in the function of the native membrane protein bacteriorhodopsin, *Soft Matter* **5**, 4899 (2009).
- [63] L. Piantanida, H. L. Bolt, N. Rozatian, S. L. Cobb, and K. Voitchovsky, Ions Modulate Stress-Induced Nanotexture in Supported Fluid Lipid Bilayers, *Biophys. J.* **113**, 426 (2017).